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ARTICLE

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Adipose-derived stem cell exosomes for treatment of dupilumab-related facial redness in patients with atopic dermatitis

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ABSTRACT

Background: Dupilumab facial redness (DFR) is a side effect of dupilumab treatment that has only been recently reported. We previously reported on two patients with DFR who were successfully treated with a topical formulation containing human adipose tissue-derived mesenchymal stem cell-derived exosomes (ASCEs).

Objectives: The study aimed to evaluate the efficacy and safety of ASCEs in DFR.

Participants and methods: We performed 12-week prospective study at single center. Twenty adult atopic dermatitis patients diagnosed with DFR were enrolled. They were treated with a topical application of the exosome formulation every week for five consecutive weeks.

Results: After exosome treatment, both the average investigator global assessment score and clinical erythema assessment scale scores decreased. 19 patients (95%) were satisfied with the treatment. Compared to baseline, erythema index at week 4 were decreased by 31, 27, 13, and 25 units on the forehead, chin, right and left cheek respectively. The analysis of stratum corneum samples revealed the expression of IL-1 α and human thymic stromal lymphopoietin was suppressed after exosome treatment, whereas filaggrin and vascular endothelial growth factor expression increased.

Conclusions: This study suggests topical formulation containing ASCEs can alleviate DFR by downregulating local inflammation and restoring skin barrier function.

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KEYWORDS

Atopic dermatitis; dupilumab; dupilumab facial redness; exosome; filaggrin; inflammation

Introduction

Dupilumab facial redness (DFR) is a recently reported side effect of dupilumab treatment for atopic dermatitis (AD). Although it was not reported as an adverse event in phase 3 clinical trials, recent studies reported DFR occurred in approximately 4–10% of patients treated with dupilumab (1–7). Erythema on visible areas including face and neck leads to a profound psychosocial burden and some patients discontinue dupilumab treatment because of this adverse event (1). Unfortunately, the etiology and treatment of this condition remain elusive (8,9).

Exosome treatment is a recently developed cell-free therapeutic strategy in regenerative medicine. Because of their small size (nanosize), biological origin, lipid bilayer membrane, ability to communicate intracellularly, and capacity to modulate the molecular activities of the recipient cell, they are attracting significant interest among dermatologists (10,11). Exosomes from mesenchymal stem cells (MSCs) are highly immunomodulatory and have shown therapeutic effects regarding wound healing, prevention of scar, skin pigmentation, and wrinkles as well as improving chronic inflammatory skin disorders such as psoriasis, AD, systemic lupus erythematosus, or bullous pemphigoid (11–23). Previously, we reported two successful cases of DFR treated with topical application of human adipose tissue-derived mesenchymal stem

cell (MSC)-derived exosomes (ASCEs) (24). In this study, we aimed to verify the efficacy of an ASCE topical formulation in treating DFR.

Materials and methods

Study design and subjects

This was a 12-week prospective study, approved by the Chung-Ang University Hospital Institutional Review Board (IRB no. 2109-014-477). Twenty Korean patients aged 18 and older, who were on dupilumab for AD and were diagnosed with DFR in Chung-Ang University Hospital (Seoul), were recruited in the study. The patients in this manuscript participated in the study voluntarily with good compliance and had given written informed consent to publication of their clinical data and photographs. The treatment and examination took 30 min to an hour.

Exosome treatment

We used the exosome formulation ASCE+SRLV-S (ExoCoBio Inc., Seoul, Republic of Korea), which contains 20 mg (970,000 ppm) of lyophilized human ASCE. Exosomes in this product were acquired from a human adipose-derived stem cell (ASC)-conditioned medium (CM) developed by ExoSCRT™ technology as described

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previously (ExoCoBio Inc., Seoul, Republic of Korea) (24,25). Briefly, ASC-CM was collected from ASCs cultured in serum-free Dulbecco's Modified Eagle's Medium (Thermo Fischer Scientific, Waltham, MA, USA), and the non-exosomal particles were removed by a 0.2 µm filter. Next, the exosomes were further purified and concentrated through tangential-flow filtration, and quantification was

performed by nanoparticle tracking analysis. Characterization of ASCEs used in this study is described in *Supplementary Figure 1*.

One vial of ASCE+SRLV-S was applied to each patient's entire face by dermatologists, and prism sonophoresis (Alummmedi, Seoul, Korea) was used to ensure effective drug delivery. This topical application of exosomes was performed weekly for five consecutive weeks.

Table 1. Rating grade for investigator global assessment score.

Grade	Description
0, clear	No inflammatory signs of atopic dermatitis
1, almost clear	Just perceptible erythema, and just perceptible papulation/infiltration
2, mild disease	Mild erythema and mild papulation/infiltration
3, moderate disease	Moderate erythema and moderate papulation/infiltration
4, severe disease	Severe erythema and severe papulation/infiltration
5, very severe disease	Severe erythema and severe papulation/infiltration with oozing /crusting

Table 2. Rating grade for clinical erythema assessment scale.

Scale	Degree of improvement
0=clear	Clear skin with no signs of erythema
1=almost clear	Almost clear, slight redness
2=mild	Definite redness, very light pink, faintly detectable erythema
3=moderate	Marked redness, dull red, clearly distinguishable erythema
4=severe	Severe erythema, fiery redness, deep, dark red

Table 3. Rating grade for patient satisfaction.

Scale	Degree of satisfaction
1	Very unsatisfied
2	Unsatisfied
3	Fair
4	Satisfied
5	Very satisfied

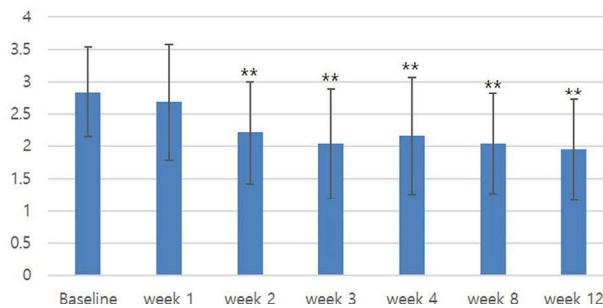
Clinical evaluation and measurement of skin parameters

Clinical and objective evaluations were performed at baseline and every week before the ASCE treatment and then at weeks 8 and 12 (four and eight weeks after the final exosome treatment). Clinical photographs were taken using the Mark-Vu® instrument (PSI plus Co. Ltd.) at each visit. The data from an investigator global assessment (IGA, 0–5, **Table 1**), a clinical erythema assessment scale (CEA, 0–4, **Table 2**), and subjective satisfaction (1–5, **Table 3**) were recorded. Skin erythema, skin hydration, and trans-epidermal water loss (TEWL) were measured using Mexameter® MX18, Corneometer® CM825, and Tewamater® TM300 (Courage & Khazaka GmbH, Cologne, Germany). Measurements were taken in accordance with the manufacturer's guidelines at the forehead, chin, right cheek, and left cheek. During the measurements, the room temperature was maintained at a constant 20–22°C and a relative humidity range of 40–60%.

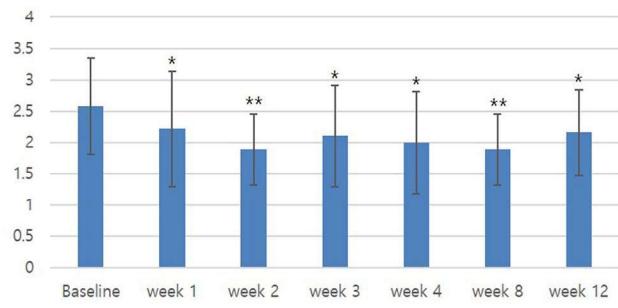
Analysis of stratum corneum samples

At baseline and week 8 (four weeks after final exosome treatment), stratum corneum samples were collected using tape stripping (skin D-squame), and subsequently analyzed in order to evaluate inflammatory cytokines and skin barrier function. Quantitative analysis of interleukin-1α (IL-1α), human thymic stromal

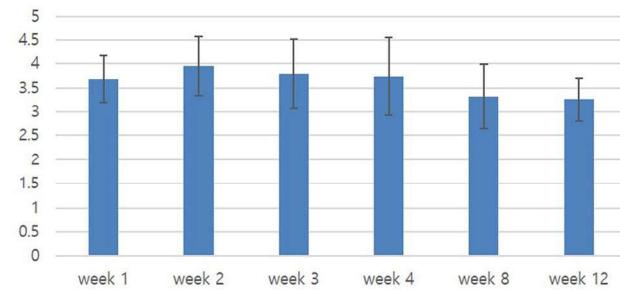
Average IGA



Average CEA



Average subject satisfaction score



*p < 0.05, ** < 0.01, *** < 0.001

Figure 1. Clinical outcomes of patients with dupilumab facial redness treated with adipose-derived stem cell exosome. Change in average investigator global assessment (IGA), clinical erythema assessment (CEA), and patient satisfaction score over time. The average IGA score decreased from week 2 and continued to decrease until week 12. The average CEA score decreased significantly from the first week after the first exosome treatment. Overall, 19 out of 20 (95%) patients had a subjective satisfaction score of 4 (satisfied) at least once during the entire clinical trial period. *p < 0.05, ** < 0.01, *** < 0.001, compared with baseline.

lymphopoietin (TSLP), filaggrin (FLG), and vascular endothelial growth factor (VEGF) was performed through real-time-polymerase chain reaction (RT-PCR) and Western blot techniques. The detailed experimental method can be found in [Supplementary Material 1](#).

Statistical analysis

We compared the resulting metabolic activities of the treatment groups and controls using one-way analysis of variance (ANOVA) and Tukey's multiple-comparison posttest. Differences between

groups were considered to significant at a p value of <0.05 . Statistical analyses were performed with GraphPad Prism 8.0 (GraphPad Software, Inc., CA, USA).

Results

Investigator global assessment, clinical erythema assessment scale, and subjective satisfaction score

The average IGA score decreased from week 2 and continued to decrease until week 12 ([Figure 1](#)). Overall, 12 out of 20 (60%)



Figure 2. Representative clinical photographs of patients. Photographs were taken at baseline, at week 2 (after the first treatment), and week 12 (eight weeks after final the treatment). After exosome treatment, clinical improvements of erythema on the both cheeks, forehead, and chin were noted.

patients achieved an IGA score of 1 (almost clear) at least once during the entire clinical trial period. Similarly, the average CEA score decreased from the first week after the first exosome treatment. A total of 11 out of 20 (55%) patients achieved a CEA score of 1 at least once during the entire clinical trial period. The average subject satisfaction score was highest in week 2 and slowly decreased over time. However, the average score was above 3 (fair) at all times. Altogether, 19 out of 20 (95%) patients had a subjective satisfaction score of 4 (satisfied) at least once during the entire clinical trial period. Representative photographs of three patients at baseline, after the first treatment, and at week 12 are shown in Figure 2.

Skin erythema, hydration, and trans-epidermal water loss

Skin erythema was measured at the forehead, chin, right cheek, and left cheek. Erythema index (EI) decreased in all four areas after exosome treatment and decreased fastest on the forehead, where it decreased from week 2. EI on the forehead was 460 at baseline, which decreased to 430 at week 2. EI decreased in all locations from week 4 (Figure 3). Compared to EI measured at baseline, EI at week 4 were decreased by 31, 27, 13, and 25 units on the forehead, chin, right and left cheek respectively. Skin

hydration and TEWL were also measured at four different anatomical areas. Although the skin hydration increased and TEWL decreased after exosome treatment, the changes were not statistically significant. (Supplementary Figures 2 and 3)

Analysis of stratum corneum samples

To determine the effect of ASCE on proinflammatory cytokine expression and epidermal proteins, we measured IL-1 α , VEGF, TSLP, and FLG expression at mRNA and protein levels (Figure 4). The mRNA expression of IL-1 α decreased in SC samples collected at week 8 compared with the baseline. By contrast, the mRNA expression of FLG and VEGF increased at week 8 compared with the baseline. TSLP was not detected at the mRNA level. Regarding protein levels, ASCE upregulated FLG expression and downregulated both IL-1 α and TSLP.

Discussion

The pathogenesis of DFR is still poorly understood, although several hypotheses exist. For example, it may represent a hypersensitivity reaction, a site-specific treatment failure, a seborrheic

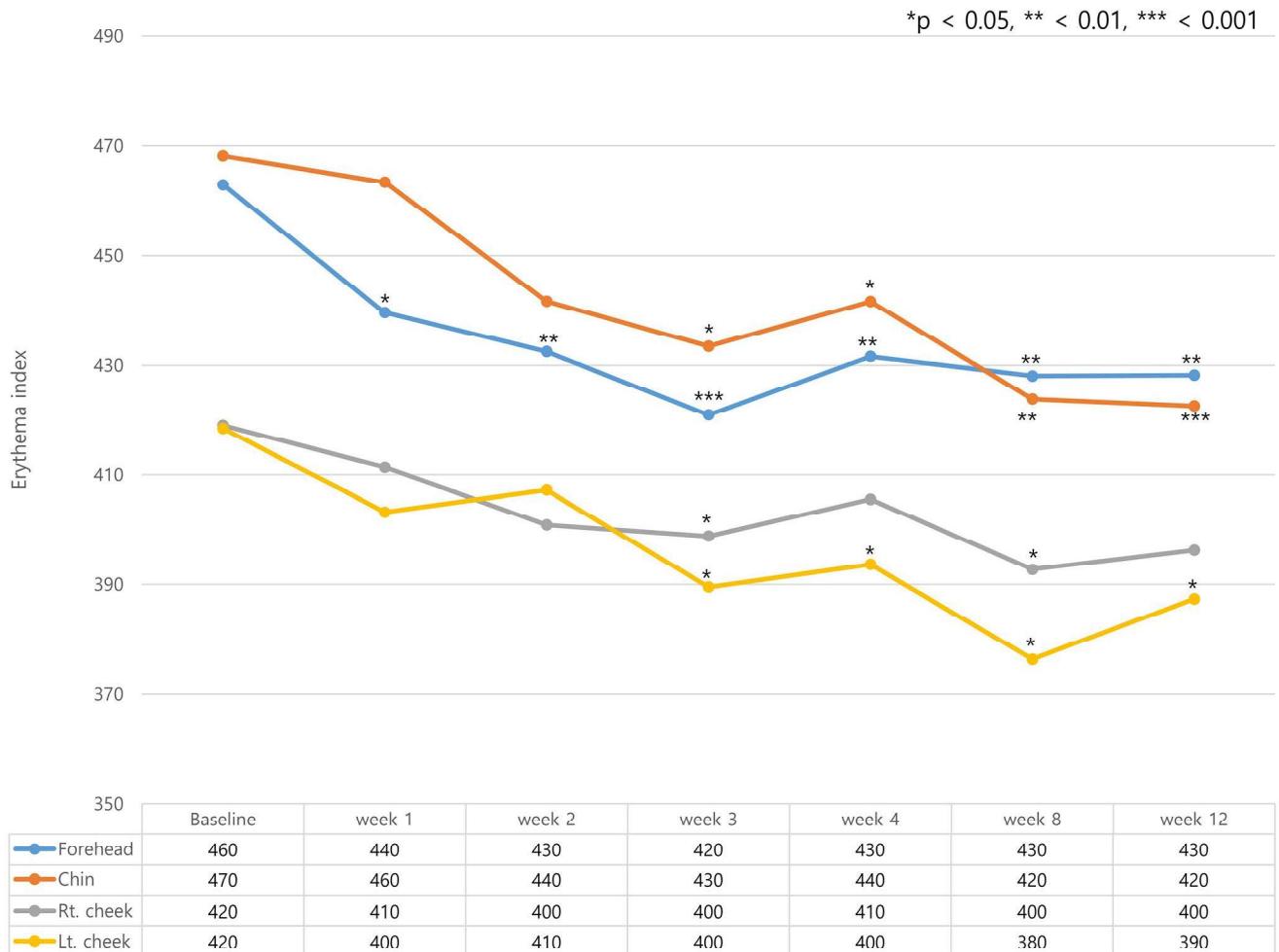


Figure 3. Changes in the erythema index (EI) were measured at four different anatomical locations on the face (forehead, left and right cheek, and chin). Erythema index (EI) decreased in all four areas after exosome treatment and decreased fastest on the forehead. *p < 0.05, ** < 0.01, *** < 0.001, compared with baseline

dermatitis-like reaction to *Malassezia*, a rosacea-like reaction associated with *Demodex*, allergic contact dermatitis, or a combination of multiple mechanisms (26). The etiology of DFR appears to vary by individual because the histologic findings also vary and include psoriasisiform dermatitis, spongiotic dermatitis with parakeratosis and neutrophil infiltration, or perifollicular and perivascular inflammation (5,27,28). For the treatment of DFR, various topical formulations including corticosteroids, calcineurin inhibitors, and antifungal agents have been attempted, but were generally unsuccessful (28–30). One potential reason for the variable treatment response could be the heterogeneous pathophysiology of DFR, meaning exosomes may be an ideal treatment option for DFR since they are highly immunomodulatory and have a pleiotropic effect on the target cells (14,18,23,31).

Our study investigated the possibility of applying a topical formulation containing ASCE to alleviate local skin inflammation seen in dupilumab-treated AD patients. Our results showed

applying topical formulation containing ASCE quickly resolves DFR symptoms. The average IGA and CEA scores decreased from the second week of treatment. Notably, CEA decreased faster than IGA, which may be because erythema improves more than other DFR symptoms such as papulation or infiltration. The clinical improvement in erythema was also supported by the decrease in EI, which is a more objective measurement. EI decreased on the forehead from week 2, and the cheeks and chin from week 4. By contrast, patient satisfaction was highest in week 2 and slowly decreased, although a score of at least 3 was maintained. This may be because ASCE treatment resulted in an immediate improvement meaning patients were satisfied with the marked change after the first few treatment sessions but thereafter became less sensitive to further improvement in their skin.

As mentioned above, studies on the successful management of DFR are very limited. The relatively few case reports of the successful treatment of DFR include one where oral minocycline,

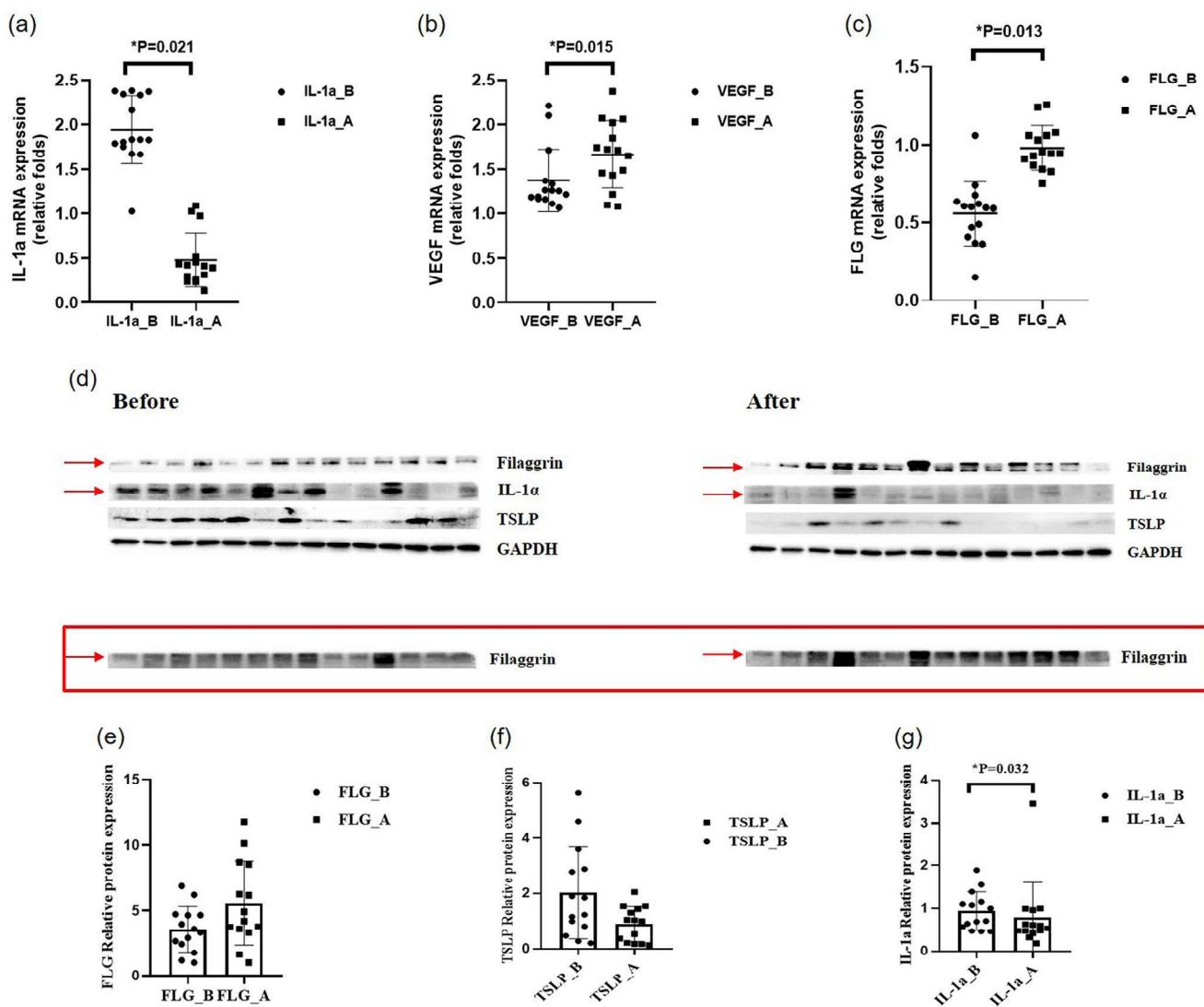


Figure 4. Results of the analysis of stratum corneum samples. (a) mRNA expression of interleukin-1 α (IL-1 α). The mRNA expression of IL-1 α decreased in SC samples collected at week 8 compared with the baseline. (b) mRNA expression of vascular endothelial growth factor (VEGF). The mRNA expression of VEGF increased in SC samples collected at week 8 compared with the baseline. (c) mRNA expression of filaggrin (FLG). The mRNA expression of FLG increased in SC samples collected at week 8 compared with the baseline. (d) Western blot analysis. (e) Protein expression of filaggrin (FLG). The protein expression of FLG increased in SC samples collected at week 8 compared with the baseline. (f) Protein expression of human thymic stromal lymphopoietin (TSLP). The protein expression of TSLP decreased in SC samples collected at week 8 compared with the baseline. (g) Protein expression of interleukin-1 α (IL-1 α). The protein expression of IL-1 α decreased in SC samples collected at week 8 compared with the baseline.

topical tacrolimus, and brimonidine tartrate gel improved facial redness within a mean of 23 weeks (5). In another case report, topical delgocitinib ointment improved DFR in one month (32). The only reported clinical study on DFR treatment is a retrospective review evaluating the efficacy of oral itraconazole. The results showed 11 of 16 (69%) DFR patients had a post-treatment IGA result of clear or almost clear (0 or 1) one to six months later, with an average self-reported improvement of 52%. Our study showed similar results, where 12 of 20 (60%) patients achieved an IGA score of 1 (almost clear), and the average subjective satisfaction score was 3.6/572. More importantly, previous treatment methods such as oral itraconazole took at least a month or more to show any therapeutic effect. By contrast, the therapeutic effect of ASCE was noticeable after just two treatment sessions.

We also observed ASCE alleviated inflammation, upregulated skin barrier-related proteins, and balanced the Th1/Th2 immune response by reducing levels of TSLP. These findings were additionally supportive of the immunomodulatory and skin barrier-restoring functions of ASCE confirmed in previous murine studies. Previous studies have reported ASCE treatment relieved AD-like symptoms and reduced inflammatory cytokines (IL-4, IL-31, IL-23, and TNF- α), and also improved epidermal barrier functions by facilitating lamellar body formation and by de-repressing the synthesis of ceramides (14,18).

In this study, we found ASCE reduced inflammatory cytokine IL-1 α . The IL-1 family cytokines initiate an inflammatory response. They have also been linked to the pathogenesis of AD as well as other inflammatory skin disorders (33,34). In particular, IL-1 α (alarmin) is released from keratinocytes in any skin condition that involves damage to the epidermis (35). Also, ASCE upregulated VEGF, which is a principal proangiogenic factor indicating ASCE can promote angiogenesis, which is consistent with many previous reports (11,19,36–39). The repair of eczema lesions not only depends on the control of inflammation, but also depends on the repair and proliferation of keratinocytes and angiogenesis (40). Last but not least, our results also demonstrated ASCE improves skin barrier function by upregulating FLG expression while suppressing TSLP (41). FLG is a key epidermal protein that contributes to skin barrier function (41,42). Additionally, TSLP plays a key role in AD pathogenesis by triggering Th2 cell inflammation and by down-regulating FLG expression (43–46). Based on these results, we can presume that topical application of ASCE effectively alleviates DFR by suppressing cytokine-mediated inflammation and restoring the skin barrier function.

The major limitation of this study is the small number of patients and the lack of a control group. Therefore, we cannot completely rule out the possibility that DFR improved spontaneously. However, based on our clinical experience and previous reports on DFR, the possibility that DFR will improve spontaneously, especially within 2 weeks, seems very small. Thus, it can be assumed that the quick clinical improvement observed in this study is largely due to the effects of ASCE treatment. Furthermore, although there are limited numbers of studies on the management of DFR, we attempted to elaborate our conclusion by comparing the magnitude of improvement observed in our study to that observed in a previous study evaluating the efficacy of oral itraconazole. Despite this, in order to make a validated conclusion, further studies that use vehicle control or that directly compares the effect of ASCE treatment with other treatment options such as oral itraconazole should be performed. In the future, larger, multi-institutional controlled studies are necessary to determine the extent of the efficacy of ASCE compared with other treatment

options, as well as to establish a proper therapeutic approach, treatment dose, and regimen.

Conclusion

As the use of dupilumab for patients with AD gradually increases, the incidence of DFR will also increase. The successful management of DFR is important in increasing patient satisfaction with their dupilumab treatment and improving their quality of life. The results of our study demonstrated the potential of a topical formulation containing ASCE in the treatment of DFR. ASCE down-regulated inflammation and increased skin barrier-related proteins and angiogenesis. Importantly, ASCE resulted in immediate clinical improvement of DFR, which has not been observed in other treatment methods. One of the most important advantages of ASCE is it can result in a therapeutic effect by topical application, which is important since DFR manifests as localized skin lesions. Thus, identifying an effective topical therapy may reduce the risks of adverse systemic effects and improve patient compliance.

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Disclosure statement

No potential conflict of interest was reported by the author(s).

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Data availability statement

The data used to support the findings of this study are available from the corresponding author upon request.

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